DUNN SCHOOL
BIOIMAGING FACILITY

FLIM
Turn ON

UNIVERSITY OF
OXFORD

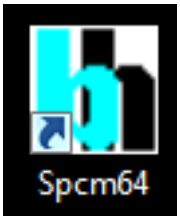
A Switch on the pulsed 488 laser

B Turn on FLIM computer

C Click on 'FLIM' remote desktop icon

D login to FLIM computer
USERNAME: microscope
PASSWORD: microscope

E Start SPC64 software



FLIM (I)

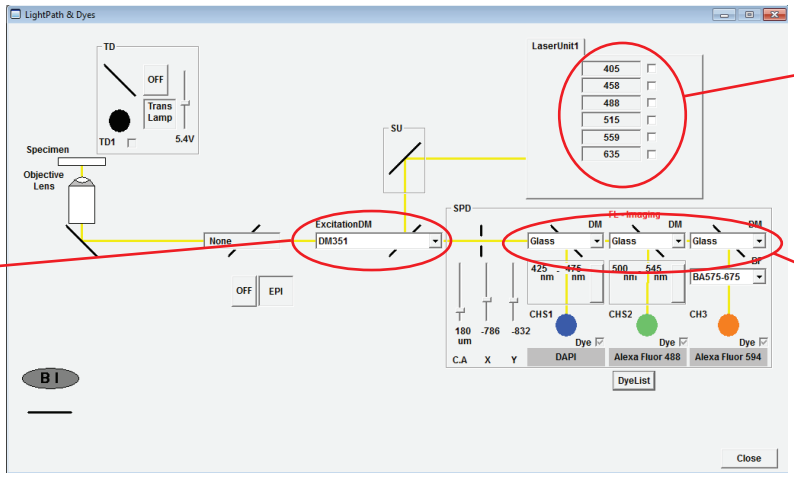
Click on Lightpath diagram icon 

Set the lightpath manually so that it looks like this:

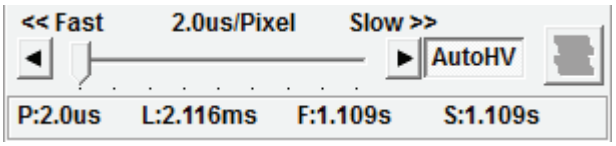
turn off all standard lasers

change Dichroic mirror to DM351

move all 3 DM to 'glass' position

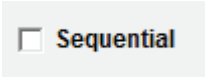


Set scan speed to 'Fast' 2.0us per pixel

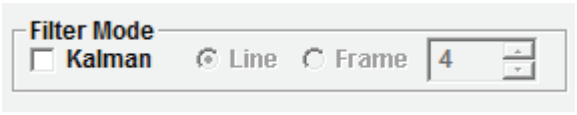


Set 'Aspect Ratio' to 512 by 512

Untick sequential scanning



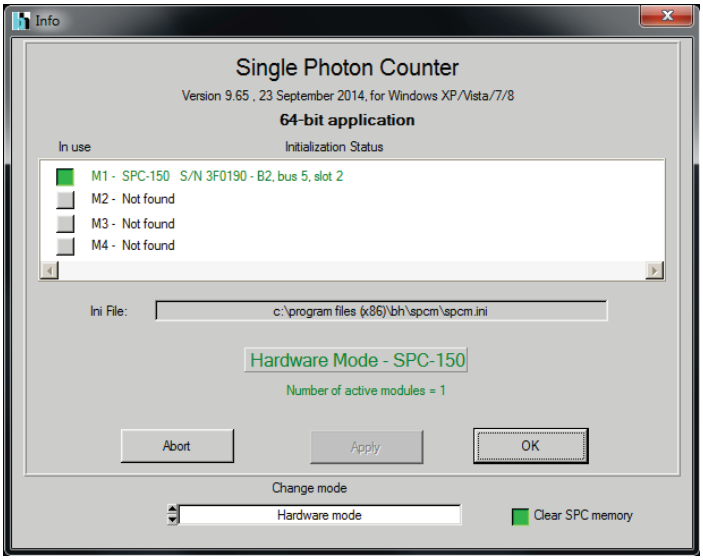
Untick 'Kalman' averaging mode



Click 'XY Repeat' scan button

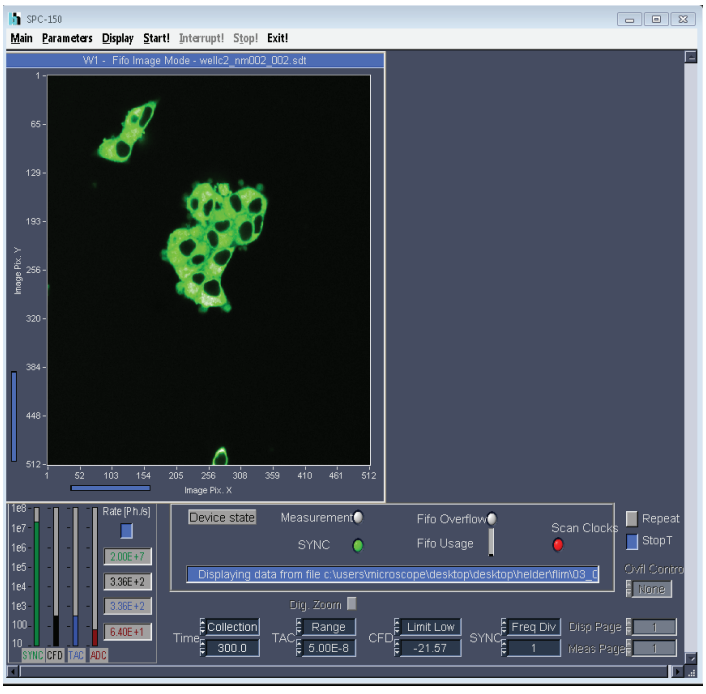
FLIM (II)

An initialization window appears
Click 'OK'



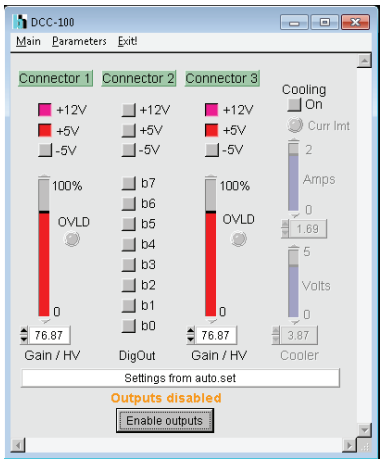
Three windows should appear:

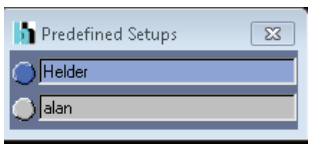
SPC-150 is the main display window



DCC-100 control the detector

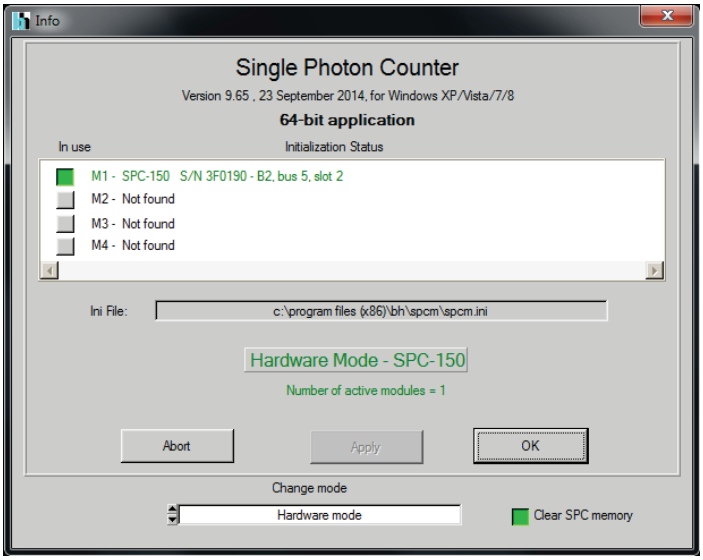
Predefined setups





FLIM (II)

An initialization window appears
Click 'OK'



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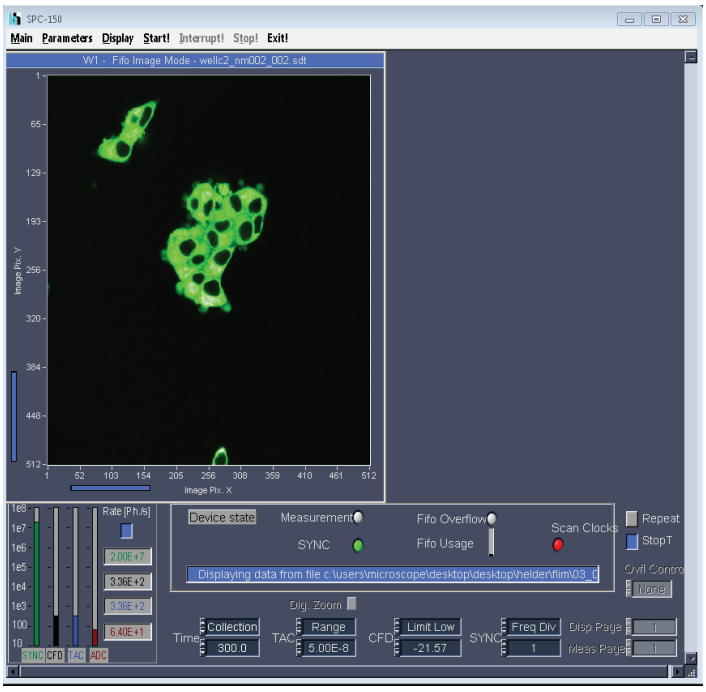
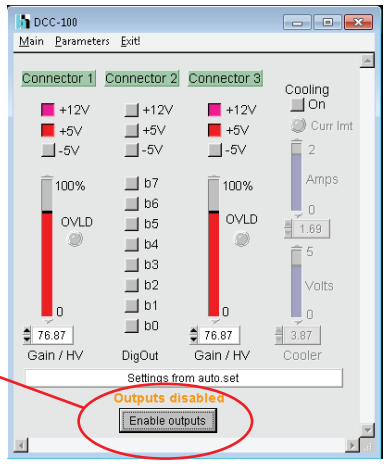
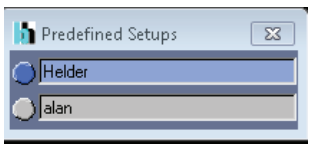
SPC-150 is the main display window



DCC-100 control the detector

Predefined setups

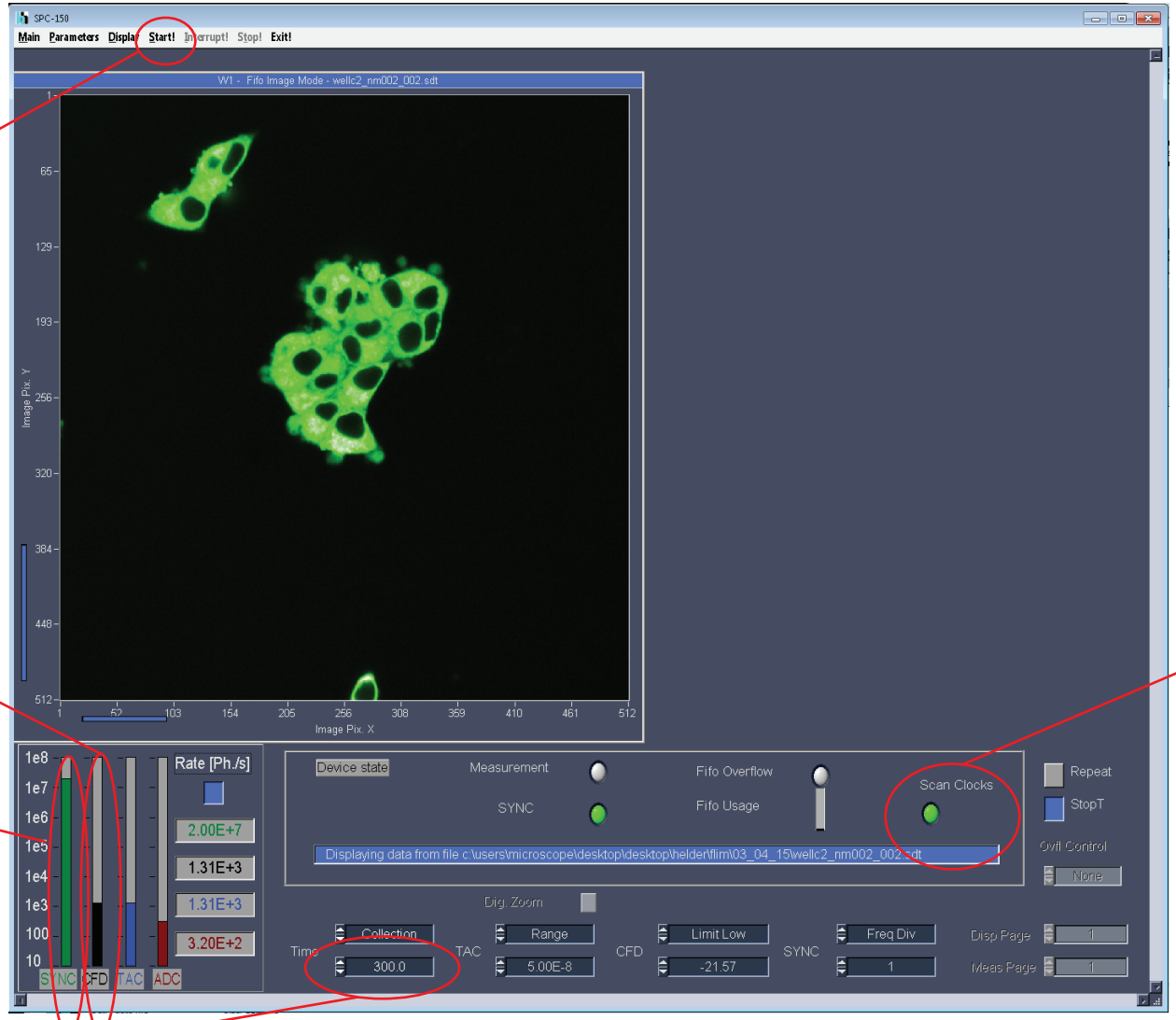




Click 'Enable outputs' to enable the detector
(the Gain / HV should be roughly 80%)

FLIM (IV)

Click 'Start!' to begin FLIM experiment



CFD indicates photon counts at the detector

SYNC indicates pulsed laser working correctly

Scan clocks green light: The FLIM computer has the correct timing from the confocal

Duration of experiment is defined here

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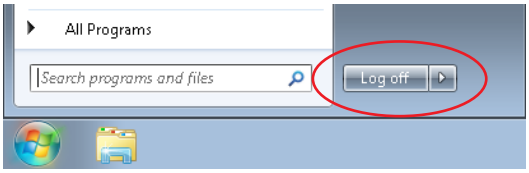
FLIM
Turn OFF

UNIVERSITY OF OXFORD

A

Close SPC64 software

Log off FLIM computer



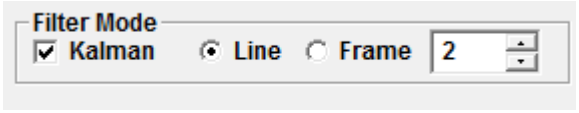
B

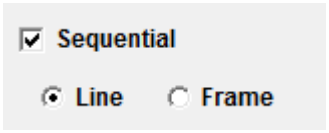
Switch off the pulsed 488 laser

C

Tick 'Kalman' averaging mode

Tick Sequential scanning





D

Return lightpath to standard 3 colour imaging
IMPORTANT - make sure last DM is in the '**MIRROR**' position.